# Improving gas exchange characteristics, antioxidant enzymes, yield and yield attributes of black cumin through foliar application of ascorbic acid

Abid Mehmood<sup>1\*</sup>, Khalid Naveed<sup>1</sup>, Muzaffar Iqbal<sup>1</sup>, Rashid Ali<sup>1</sup>, Malik Faizan Shaukat<sup>2</sup>, Haris Shahzad<sup>2</sup>, Anees Ur Rehman<sup>1</sup>, Ahmad Ayub<sup>2</sup>, Mubasir Ahmed<sup>1</sup>, Diyan Ahmad<sup>1</sup>, Imad Ali<sup>1</sup>, Abdul Tawab<sup>1</sup>, Saifur Rehman<sup>1</sup>, Saad Hasan<sup>3</sup>, Sagheer Khan<sup>4</sup>, Qammer Shahzad<sup>4</sup>, Nauman Shehzad<sup>3</sup>, Riaz Hussain<sup>1</sup>, Muhammad Umer Khatab<sup>3</sup> and Naveed Ul Haq<sup>4,5</sup>

- 1. Department of Agronomy, The University of Haripur, Pakistan
- 2. Department of Horticulture, The University of Haripur, Pakistan
- 3. Department of Soil & Climate Science, The University of Haripur, Pakistan
- 4. Department of Food Science and Technology, The University of Haripur, Pakistan
- 5. Department of Food Science, The University of Guelph, Canada
- \* Corresponding author's email: abidawan1990@gmail.com

# Abstract

Plant growth regulators are known to improve plant's vegetative growth and yield when applied at very low concentrations. Ascorbic acid (Vitamin-C) plays a vital role in sustaining plant growth among the numerous plant growth regulators that control growth under normal or stress conditions. An experiment was conducted during rabi 2018-19 to assess the performance of black cumin crop when sprayed with different ascorbic acid concentrations, with the objective of digging out the best ascorbic acid concentration for enhancing the production of black cumin under climatic conditions of Haripur. The results showed that 100  $\mu$ M ascorbic acid was found to be effective followed by 150 $\mu$ M ascorbic acid. Photosynthetic rate (m mol (CO<sub>2</sub>) m<sup>-2</sup> s<sup>-1</sup>), Stomatal conductance (mmol H<sub>2</sub>O m<sup>-2</sup> s<sup>-1</sup>), Net CO<sub>2</sub> assimilation rate ( $\mu$ mol CO<sub>2</sub> m<sup>-2</sup> s<sup>-1</sup>), Transpiration rate (mmol H<sub>2</sub>O m<sup>-1</sup> s<sup>-1</sup>), Peroxidase (POX), Plant height, number of capsules plant<sup>-1</sup>, seeds capsule<sup>-1</sup>, 1000-seed weight (g) and Seed yield (kg ha<sup>-1</sup>) were significantly affected from ascorbic acid concentrations. However, as this study was conducted with only with ascorbic acid, it is advisable to conduct an experiment with various other growth regulators, their combinations and different concentrations to check out their synergistic effects on production traits of black cumin to make reliable and acceptable recommendations.

**Keywords**: Black cumin, Photosynthesis, antioxidant enzymes, growth regulator, ascorbic acid **Introduction**:

The production of medicinal plants become ultimate goal because of the demand for medicinal plants around the world (Shahzad et al., 2020). However, these crops need special attention, using suitable and recommended agronomic practices to obtain maximum income as compared with traditional crops. Black cumin, Nigella sativa L. commonly known as kalonji (Rehman et al., 2022) is one of the most widely examined plant possessing naturally occurring compounds with anti-cancer potential. The numerous vernacular names indicate the use of the spice in more than 100 countries (Mehmood *et al.*, 2021a). The seeds are bitter in taste and consumption of whole seed even in small quantity gives a feeling of constriction of throat (Saul *et al.*, 2013). The seeds of black cumin crop contain approximately 30- 35 % of oil which can be utilized in food and pharmaceutical industries (Mehmood et al., 2018). The production of black cumin in Pakistan has not been reported on commercial scale but it is widely used for various purposes in the country. With increase in demand for medicinal plants, black cumin is a potential specie for crop diversification and it can also reduce the risk of crop failure from which economic condition of smaller land holdings could also be improved (Hussain et al., 2003: Mehmood et al., 2021b). The major crops grown in Haripur are wheat, maize and peas thus offering limited choice of seed spice crops for the farmers. Introduction of new seed spices open new avenues for the farmers by not only widening the choice of crops but also improve their revenue from the land (Ayub et al., 2021). In such a scenario, among the several suitable seed spice crops, black cumin demands special attention due to its export value as well as increasing demand as medicinal spice (Khan et al., 2020).

Yield is dependent to a large extent on soil nutrient status and external application of nutrients (Ayub et al., 2020). Beside the importance of synthetic fertilizers, they have many adverse effects environment, human and animals' health, loss of soil fertility and also their cost is very high higher (Boraste *et al.*, 2009; Mehmood et al., 2020). Finding the alternate methods for supplying essential nutrients to the crop is need of the hour (Hassan and Ali, 2013). The physiological efficiency of plants is affected by plant growth regulators (Solaimalai *et al.*, 2001). They influence growth and yield of plants when applied at a very low concentration (Naeem *et al.*, 2004; Ayuba et al., 2020). In addition, the impact of plant growth regulators on crops may

#### Journal of Xi'an Shiyou University, Natural Science Edition

differ with the type of plants, their genetic characteristics, variety, stage of development, environmental conditions, physiological and nutritional status, and senescence delays in endogenous hormonal equilibrium (Adebayo *et al.*, 2011).

Ascorbic acid (Vitamin C) is synthesized and affects the growth and production of plants in higher plants and plays an important part in the electron transport mechanism Blokhina *et al.*, (2003) reported that Ascorbic acid has potential as regulator of cell division and differentiation and to play a significant role in a broad range of functions, such as antioxidant defense, photosynthesis control and development (Allahveran *et al.*, 2018). Ascorbic acid is a cofactor for certain enzymes, such as those active in the synthesis of the cell wall and proline residue hydroxylation (Abdel-Hafeez *et al.*, 2019).

However, little information seems to be available on the concentration of ascorbic acid for foliar application on black cumin crop to enhance growth and obtain maximum yield. Keeping this in mind, present investigation was performed in order to identify the best ascorbic acid concentration for maximizing yield and its components of black cumin under Haripur conditions.

# Materials and methods:

# Site description:

The present research was carried out at the Agricultural Farm, Department of Agronomy, The University of Haripur, KP during rabi season 2018-19. This study was aimed find photosynthetic efficiency, antioxidant enzymes and yield and yield attributes of black cumin through foliar application of ascorbic acid under climatic conditions of Haripur.

Soil samples from the depth of 0 and 15 cm were collected and a composite sample was prepared for soil analysis before preparatory cultivation. Standard methods were followed for physiochemical properties of soil (Table 1).

#### **Experimentation:**

RCBD was used for the arrangement of experiment. Each treatment was replicated three times. Black cumin variety NARC-1 kalonji was used. Seeds were sown on first week of November. Seeds were sown in rows having 30 cm row spacing. The plot size was  $2 \times 3 \text{ m}^2$ . NPK was applied at the rate of 40-20-20.

Ascorbic acid was applied as foliar application as T1= 0 $\mu$ M, T2= 50 $\mu$ M, T3= 100 $\mu$ M, T4= 150 $\mu$ M, T5= 200 $\mu$ M, T6= 250 $\mu$ M, T7= 300 $\mu$ M. These were applied 90 days after sowing. 0  $\mu$ M ascorbic acid plots were sprayed with distilled water only.

#### Journal of Xi'an Shiyou University, Natural Science Edition

Data on Physiological characters like photosynthetic rate, transpiration rate, stomatal conductance, transpiration rate and Net CO<sub>2</sub> assimilation rate, were recorded on youngest leaf which are fully expended from each plant with the help of an open system LCA-4 ADC portable Infrared gas analyzer (Analytical Development Company, Hoddesdon, England). The readings were recorded from 10:15 to 11:45 with the following specifications/adjustments: leaf surface area 3.25 cm<sup>2</sup>, temperature of leaf chamber varied from 31.5 to 37.8 C, leaf chamber gas flow rate (v) 392.8 ml min<sup>-1</sup>, Molar flow of air per unit leaf area (Us) 404.84 mol m<sup>-2</sup>sec<sup>-1</sup>. Ambient chamber ranged from 20.5–23.1 m bar, PAR (Qleaf) at leaf surface was maximum up to 1048 1 mol m<sup>-2</sup>s<sup>-</sup>1. The peroxidase (POX), catalase (CAT), and ascorbate peroxidase (APX) behaviours were spectrophotometrically determined. Leaves were homogenised as defined by Dixit *et al.*, (2001) in a medium composed of 50 mM phosphate buffer with 7.0 pH and 1 mM dithiothreitol (DTT). Plant height, capsules plant<sup>-1</sup>, capsule<sup>-1</sup> seeds, 1000-grain weight (g) and seed yield (kg ha<sup>-1</sup>) were recorded. The analysis of variance (ANOVA) was worked out from collected data to observe difference among various treatments (Gomez and Gomez, 1984).

Texture	PH	%	Organic	Ν	Р	K	Ca <sup>+</sup>	$Mg^+$	HCO <sub>3</sub>	Cl
class	value	TSS	matter	(%)	(ppm)	(ppm)				
Silty	7.41	1.2	1.71	0.089	9.54	68	5.00	0.95	5.68	0.11
Clay										

# **RESULTS AND DISCUSSIONS**

# Photosynthetic rate (mmol (CO<sub>2</sub>) m<sup>-2</sup> s<sup>-1</sup>):

Photosynthetic rate of black cumin was significantly affected from ascorbic acid concentrations (table 2). Statistically similar results regarding highest photosynthetic rate was recorded in those plants sprayed with ascorbic acid @ 150  $\mu$ M (15.5 (mmol (CO<sub>2</sub>) m<sup>-2</sup> s<sup>-1</sup>) and plants treated with ascorbic acid concentration of 100  $\mu$ M (15.2(mmol (CO<sub>2</sub>) m<sup>-2</sup> s<sup>-1</sup>), followed by (13.8(mmol (CO<sub>2</sub>) m<sup>-2</sup> s<sup>-1</sup>)) the plants sprayed with ascorbic acid @ 50  $\mu$ M, while lowest photosynthetic rate (8.2(mmol (CO<sub>2</sub>) m<sup>-2</sup> s<sup>-1</sup>) was observed in control treatment (table 03). In photosynthesis, ascorbic acid has a central position, as the high concentration in chloroplasts would suggest. In its three biochemical modes, it functions. Firstly, by removing hydrogen peroxide produced by oxygen photo reduction, it acts as an antioxidant (Akram *et al.*, 2017). This series has been

#### Journal of Xi'an Shiyou University, Natural Science Edition

dubbed the reaction of Mehler peroxidase (Aziz *et al.*, 2018). Secondly, mono-dehydro ascorbate, formed by ascorbate peroxidase can act as a direct electron acceptor to PSI (Hussein and Mohamed, 2017). Thirdly, it is a violaxanthin de-epoxidase cofactor. In bright sun, or when low temperatures and drought limit carbon dioxide fixation, zeaxanthin dissipates the excess excitation energy as heat in the light harvesting antennae (Mohamed *et al.*, 2017).

 Table 02: Analysis of variance results (Means squares) of various traits of black cumin as affected from ascorbic acid concentrations

Source	DF	Photosynthetic	Stomatal	Net CO <sub>2</sub>	Transpiration	CAT	POX
		rate	Conductance	assimilation	rate		
				rate			
Replication	2	0.1300	0.00010	0.08020	0.01857	63.4762	92.33
Treatment	6	21.5997	0.00489	3.07004	2.03857	82.8889 <sup>NS</sup>	4344.41
Error	12	0.5228	0.00011	0.09592	0.08690	62.1984	54.56
CV %		5.81	3.21	3.56	6.92	1.37	0.34
Mean		12.443	0.3286	8.704	4.2571	574.67	2162.2

Significant at 0.05 level, NS= Non-Significant

# Stomatal conductance (mmol H<sub>2</sub>O m<sup>-2</sup> s<sup>-1</sup>):

Table 02 depicting analysis of variance of stomatal conductance of black cumin affected from various concentrations of ascorbic acid. Significant differences were observed regarding foliar application of various ascorbic acid concentrations. Statistically similar results regarding highest stomatal conductance was noted in the application of ascorbic acid at concentration of 100  $\mu$ M (0.38 mmol H<sub>2</sub>O m<sup>-2</sup> s<sup>-1</sup>) and 150  $\mu$ M (0.37 mmol H<sub>2</sub>O m<sup>-2</sup> s<sup>-1</sup>). While statistically lower stomatal conductance was observed 300  $\mu$ M (0.28 mmol H<sub>2</sub>O m<sup>-2</sup> s<sup>-1</sup>) and in control treatment (0.27 mmol H<sub>2</sub>O m<sup>-2</sup> s<sup>-1</sup>). To reduce transpiration rate of leaf and to maintain turgor of cell for maintaining normal biochemical processes, regulation of stomatal conductance is an initial defense mechanism of plants to various stresses (Mehmood *et al.*, 2021). Studies have shown that ascorbic acid has an effect on stomatal conductance in barley (Habibi, 2013), chickpea (Rahbarian *et al.*, 2011), wheat (Moud and Yamagishi, 2005), and in potato (Kawakami *et al.*, 2006).

Net CO<sub>2</sub> assimilation rate (µmol CO<sub>2</sub> m<sup>-2</sup> s<sup>-1</sup>):

Net CO<sub>2</sub> assimilation rate was affected significantly from different concentrations of ascorbic acid (table 02). Maximum CO<sub>2</sub> assimilation rate (10.3 µmol CO<sub>2</sub> m<sup>-2</sup> s<sup>-1</sup>) was observed in plants sprayed with 100 µM, followed by (9.3 µmol CO<sub>2</sub> m<sup>-2</sup> s<sup>-1</sup>) plants sprayed with 150 µM. whereas, minimum CO<sub>2</sub> assimilation rate (7.3 µmol CO<sub>2</sub> m<sup>-2</sup> s<sup>-1</sup>) was observed in crop plots. Ascorbic acid is considered to have a key role in photosynthesis, stomatal conductance and the net rate of CO<sub>2</sub> assimilation is characterized by its occurrence in all photosynthetic tissues where reversed hydration of CO<sub>2</sub> is catalyzed. In the present study, plants treated with medium ascorbic acid concentration have possessed greater CO<sub>2</sub> assimilation rate as compared to the control plants. This effect is likely caused by influence of ascorbic acid on the CO<sub>2</sub> assimilation rate, which involves translation/transcription (Alam *et al.*, 2012).

Table 03: Means of photosynthetic rate, stomatal conductance, transpiration rate, CO<sub>2</sub> assimilation rate, POX and CAT of black cumin as affected from ascorbic acid concentrations

	Photosynthetic	Stomatal	Net CO <sub>2</sub>	Transpiration	CAT	POX
	rate	Conductance	assimilation	rate		
			rate			
<b>T1</b>	8.2 e	0.27 d	7.3 d	2.7 e	574.3 <sup>NS</sup>	2096.3 f
T2	13.8 b	0.34 b	9.0 b	4.5 bc	577.3	2193.3 b
Т3	15.2 a	0.38 a	10.3 a	5.1 a	581.0	2211.3 a
<b>T4</b>	15.5 a	0.37 a	9.3 b	4.9 ab	577.3	2179.3 c
T5	12.6 bc	0.34 b	8.8 b	4.4 bc	577.0	2156 d
<b>T6</b>	11.5 c	0.31 c	8.2 c	4.2 cd	565.3	2162.3 d
<b>T7</b>	10.2 d	0.28 d	7.8 с	3.7 d	570.3	2137 e
LSD	1.28	0.01	0.55	0.52	14.03	13.14

NS= Non-significant

Transpiration rate (mmol H<sub>2</sub>O m<sup>-1</sup> s<sup>-1</sup>):

Transpiration rate (mmol H<sub>2</sub>O m<sup>-1</sup> s<sup>-1</sup>) was affected significantly from different concentrations of ascorbic acid (table 02). Highest Transpiration rate (5.1 mmol H<sub>2</sub>O m<sup>-1</sup> s<sup>-1</sup>) was observed in plants sprayed with 100  $\mu$ M, followed by (4.9 mmol H<sub>2</sub>O m<sup>-1</sup> s<sup>-1</sup>) plants sprayed with 150  $\mu$ M. whereas, minimum transpiration rate (2.7 mmol H<sub>2</sub>O m<sup>-1</sup> s<sup>-1</sup>) was observed in crop plots. Plants sprayed with ascorbic acid exhibited higher transpiration rate than the control. Medlyn *et al.*, (2013) proposed that function of stomata may be governed by mesophyll activities; hence, reduced transpiration rate facilitates the free exchange of gases from plants.

# Catalase (CAT):

Table 02 depicting analysis of variance results of catalase activity of black cumin as affected from various ascorbic acid concentrations. Results regarding catalase activity showed non-significant differences. Highest CAT was seen in plants sprayed with ascorbic acid @ 100  $\mu$ M while lowest was noted in plants sprayed with ascorbic acid @ 250  $\mu$ M.

# **Peroxidase (POX):**

Analysis of variance for peroxidase showed significant differences with various concentrations of ascorbic acid application (table 02). Highest POX (2211.3 min<sup>-1</sup> g<sup>-1</sup> FW) was observed in ascorbic acid concentration of 100  $\mu$ M, followed by (2193.3 min<sup>-1</sup> g<sup>-1</sup> FW) plants sprayed with ascorbic acid @ 50  $\mu$ M, whereas, lowest POX value (2096.3 min<sup>-1</sup> g<sup>-1</sup> FW) was observed in control treatment. Increased POX activation / levels were reported by Khan *et al.*, 2012 in the *Pteris vittata* L ascorbic acid accumulator. Using the incorporation of ascorbic acids exposed to different stresses, for example in case of rapeseed seedlings when subjected to drought stress (Hasanuzzaman and Fujita, 2014), Phytotoxicity in wheat seedlings under cold stress and in barley under ascorbic acid, which could be directly related to oxidative stress caused by production of ROS, i.e. H<sub>2</sub>O<sub>2</sub> and superoxide anion O<sub>2</sub> or, indirectly, by decreasing the content of glutathione (Benavides *et al.*, 2005).

# Plant height (cm):

Plant height of black cumin was significantly affected from ascorbic acid concentrations (Table 04). Tallest plants (86.63 cm) were recorded with foliar application of 150  $\mu$ M ascorbic acid concentration followed by 83.9 cm when ascorbic acid was sprayed @ 200  $\mu$ M. Whereas, shortest plants were observed in plots sprayed with distilled water only (64.3 cm) (table 05). Plant growth regulators are chemicals which influence plant growth and yield when applied at a

very low concentration. It is very well known that major development processes in plants are controlled by various group of plant hormones which sometimes act as growth promotor when applied in lower concentrations while in higher concentrations they act as growth retarders. It was thus apparent that these two treatments among other treatments caused maximum plant height. ascorbic acid is known to be a significant plant growth regulator that has been documented to promote a broad variety of processes, including cell elongation, contributing to increased development and growth (Sadak *et al.*, 2015).

#### Capsules plant<sup>-1</sup>:

Significant difference among various ascorbic acid concentrations was observed during the study (table 04). Maximum number of pods/capsules plant<sup>-1</sup> (43.6) was produced by the plants sprayed with 100  $\mu$ M ascorbic acid followed by (40.66) when ascorbic acid was applied @ 150  $\mu$ M. Whereas, minimum capsules plant<sup>-1</sup> (28.6) were recorded in those plants which were not treated with growth hormones (Table 05). It is interesting to note here that lower concentration of ascorbic acid (100  $\mu$ M) increased the capsules number plant<sup>-1</sup> than higher concentrations. Maximized photosynthesis and improved sink capability can be related to these important results by providing photo-assimilates from leaves and translocation to create high-quality fruit and yield as reported by Thomas and Howarth, 2000 also revealed that the application of growth hormones induced bio-synthesis of cytokinin in turned maximum number of active photosynthetic leaves and that is apparent from the number and area of leaves per plant retaining higher concentrations of chlorophylls reflecting on plant yield.

Table 04: Analysis of variance results (Means squares) of various yield related traits of
black cumin as affected from ascorbic acid concentrations

Source	DF	Plant height	Capsules	Seeds	1000-seed	Seed yield
			plant <sup>-1</sup>	capsule <sup>-1</sup>	weight	kg ha <sup>-1</sup>
Replication	2	20.195	8.1905	28.5962	0.00123	687.83
Treatment	6	202.863	94.6349	58.7638	0.19682	6679.89
Error	12	1.582	1.3016	5.9962	0.00458	224.87
CV %		1.68	3.28	2.63	2.61	2.73
Mean		74.952	34.762	93.195	2.5910	549.21

Significant at 0.05 level

# Seeds capsule<sup>-1</sup>:

Table 04 showed that all levels of ascorbic acid lead to significant increases in seeds capsule<sup>-1</sup> as compared to the control treatment. Data also cleared that spraying ascorbic acid @ 100  $\mu$ M on black cumin plants was sufficient to produce the superiority results of seeds capsule<sup>-1</sup> as compared to all other concentrations. ascorbic acid foliar application with low concentration provides adequate quantities of stimulant substances promoted to increase the rate of cell division, cell enlargement, growing plants, gradually generating more and higher seed. These results are in full agreement with those obtained by Kandil *et al.*, (2007) who found that exogenously applied ascorbic acid 100 ppm maximize seed yield and its components due to reducing flower, shedding percentage, lodging of plants and an escalating yield component.

 Table 05: Effect of different concentrations of ascorbic acid on growth and yield of Black

 cumin

	Plant height	Number of	Number of	1000-seed	Seed yield kg ha <sup>-1</sup>
		capsules	seeds	weight	
		plant <sup>-1</sup>	capsule <sup>-1</sup>		
T1	64.3 f	28.66 e	86.8 e	2.25 e	488.89 e
T2	72.36 d	36.33 c	89.1 de	2.30 e	500.00 de
T3	66.7 c	43.66 a	97.2 bc	2.91 a	605.5 a
T4	86.63 a	40.66 b	99.4 a	2.73 bc	577.7 b
Т5	83.93 b	31.66 d	94.6 bc	2.82 ab	600.00 ab
<b>T6</b>	75.26 c	31.00 d	93.6 bc	2.63 c	522.22 d
Т7	75.46 c	31.33 d	91.5 cd	2.46 d	550.00 c
LSD	2.23	2.02	4.35	0.12	26.67

1000-seed weight:

1000-seed weight of black cumin was affected significantly from various ascorbic acid concentrations. Application of ascorbic acid @ 100  $\mu$ M showed promising results and was significantly on par, followed by ascorbic acid concentration of 150  $\mu$ M. While least 1000-seed weight were observed in plots sprayed with distilled water only. Chattha *et al.* (2015) also found the increased 1000-grain weight due to application of growth enhancing substances applied at different concentrations.

# Seed yield kg ha<sup>-1</sup>:

All concentrations of ascorbic acid improve seed yield. However, highest seed yield (605.5 kg ha<sup>-1</sup>) was recorded with 100  $\mu$ M ascorbic acid, followed by (600 kg ha<sup>-1</sup>) 200  $\mu$ M ascorbic acid, whereas, lowest seed yield was observed in control plots (488.8 kg ha<sup>-1</sup>). The increase in seed yield may be due to the defensive effect of ascorbic acid in increasing assimilates and their translocation from leaves to fruits where the weight of the seeds increased (Ibrahim *et al.*, 2007).

# **Conclusion:**

Form this study it can be concluded that ascorbic acid @  $100 \ \mu M$  was more effective for enhancing the growth, yield and yield components of black cumin under the climatic conditions of Haripur Hazara, Khyber Pakhtunkhwa, Pakistan.

# **REFERENCES:**

- Abdel-Hafeez, A. A., Abd El-Mageed, T. A., & Rady, M. M. (2019). Impact of ascorbic acid foliar spray and seed treatment with cyanobacteria on growth and yield component of sunflower plants under saline soil conditions. *International Letters of Natural Sciences*, 76, 136-146.
- Adebayo, A.G., Akintoye, H.A., Olufolaji, A.O., Aina, O.O., Olatunji, M.T. and Shokalu, A.O., 2011. Assessment of organic amendments on vegetative development and nutrient uptake of Moringa oleifera Lam in the nursery. *Asian Journal of Plant Sciences*, 10(1), p.74.
- Akram, N. A., Shafiq, F., & Ashraf, M. (2017). Ascorbic acid-a potential oxidant scavenger and its role in plant development and abiotic stress tolerance. *Frontiers in plant science*, 8, 613.

- Alam, M. M., Naeem, M., Idrees, M., Masroor, M., & Khan, A. (2012). Augmentation of photosynthesis, crop productivity, enzyme activities and alkaloids production in Sadabahar (*Catharanthus roseus* L.) through application of diverse plant growth regulators. *Journal of Crop Science and Biotechnology*, 15(2), 117-129.
- Allahveran, A., Farokhzad, A., Asghari, M. and Sarkhosh, A., 2018. Foliar application of ascorbic and citric acids enhanced 'Red Spur'apple fruit quality, bioactive compounds and antioxidant activity. *Physiology and Molecular Biology of Plants*, 24(3), pp.433-440.
- Ayub, Q., Khan, S.M., Hussain, I., Gurmani, A.R., Naveed, K., Mehmood, A., Ali, S., Ahmad, T., Haq, N.U. and Hussain, A., 2021. Mitigating the adverse effects of NaCl salinity on pod yield and ionic attributes of okra plants by silicon and gibberellic acid application. *Italus Hortus*, 28, p.59. <u>https://doi.org/10.26353/j.itahort/2021.1.5973</u>
- Ayub, Q., Khan, S.M., Hussain, I., Naveed, K., Ali, S., Mehmood, A., Khan, M.J., Haq, N.U. and Shehzad, Q., 2021. Responses of different okra (Abelmoschus esculentus) cultivars to water deficit conditions. *Journal of Horticultural Sciences*, 16(1), pp.53-63. https://jhs.iihr.res.in/index.php/jhs/article/view/1099
- Ayub, Q., Mehmood, A., Hayat, U., Shahzad, Q. and Ahmad, S., 2020. 7. Effect of salinity on physiological and biochemical attributes of different Brinjal (Solanum melongena L.) cultivars. *Pure and Applied Biology (PAB)*, 9(4), pp.2190-2198.
  <u>http://dx.doi.org/10.19045/bspab.2020.90234</u>
- Ayuba, Q., Khana, S.M., Mehmoodb, A., Haqc, N.U., Alia, S., Ahmadd, T., Ayuba, M.U., Hassaana, M., Hayata, U. and Shoukata, M.F., 2020. Enhancement of Physiological and Biochemical Attributes of Okra by Application of Salicylic Acid under Drought Stress. *Journal of Horticultural Science and Technology*, 3(4), pp.113-119. <u>https://doi.org/10.46653/jhst2034113</u>
- Aziz, A., Akram, N. A., & Ashraf, M. (2018). Influence of natural and synthetic vitamin C (ascorbic acid) on primary and secondary metabolites and associated metabolism in quinoa (*Chenopodium quinoa*) plants under water deficit regimes. *Plant Physiology and Biochemistry*, 123, 192-203.
- Benavides, M. P., Gallego, S. M., & Tomaro, M. L. (2005). Cadmium toxicity in plants. *Brazilian journal of plant physiology*, *17*(1), 21-34.

- Boraste, A., Vamsi, K. K., Jhadav, A., Khairnar, Y., Gupta, N., Trivedi, S., ... & Joshi, B. (2009). Biofertilizers: A novel tool for agriculture. *International Journal of Microbiology Research*, 1(2), 23.
- Chattha, M. U., Sana, M. A., Munir, H., Ashraf, U., Zamir, S. I., & Ul-Haq, I. (2015). Exogenous application of plant growth promoting substances enhances the growth, yield and quality of maize ('Zea mays' L.). Plant Knowledge Journal, 4(1), 1.
- Dixit, V., Pandey, V., & Shyam, R. (2001). Differential antioxidative responses to cadmium in roots and leaves of pea (*Pisum sativum L.*). *Journal of Experimental Botany*, 52(358), 1101-1109.
- Gomez, K. A., & Gomez, A. A. (1984). Statistical procedures for agricultural research. John Wiley & Sons.
- Habibi, G. (2013). Effect of drought stress and selenium spraying on photosynthesis and antioxidant activity of spring barle. *Acta Agriculturae Slovenica*, *101*(1), 31.
- Hasanuzzaman, M., Nahar, K., Alam, M. M., & Fujita, M. (2014). Modulation of antioxidant machinery and the methylglyoxal detoxification system in selenium-supplemented Brassica napus seedlings confers tolerance to high temperature stress. *Biological trace element research*, 161(3), 297-307.
- Hassan, F. A. S., & Ali, E. F. (2013). A Comparative Study between Traditional Mineral Nutrition and Alternative Sources on Anise Plant. *European Journal of Scientific Research*, 106(2), 201-212.
- Hussain, K., Rabbi, F., Akbar, U., & Rafiq, M. (2003). Effect of different concentrations of indole acetic acid (IAA) on growth and yield of black seeds. Sarhad Journal of Agriculture (Pakistan).
- Hussein, H. M., & Mohamed, A. H. (2017) Growth, Flowering, Yield and Oil Characteristics of Picual Olives as Affected by Foliar Application of Vitamins and Amino Acids.*New York Science Journal.* 10(7), pp. 79-85.
- Ibrahim, M. E., Bekheta, M. A., El-Moursi, A., & Gaafar, N. A. (2007). Improvement of growth and seed yield quality of *Vicia faba* L. plants as affected by application of some bioregulators. *Australian Journal of Basic and Applied Sciences*, 1(4), 657-666.

- Kandil, M. M., Shalaby, M. A., & Mahgoub, M. H. (2007). Effect of some growth regulators on the levels of endogenous hormones chemical and constituents of rose plant. *American-Eurasian J. Agtic. Environ. Sci*, 2, 720-730.
- Kawakami, J., Iwama, K., & Jitsuyama, Y. (2006). Soil water stress and the growth and yield of potato plants grown from microtubers and conventional seed tubers. *Field Crops Research*, 95(1), 89-96.
- Khan, M.J., Ayub, Q., Hussain, I., Mehmood, A., Arif, N., Mehmood, S., Shehzad, Q., Khalid, S. and Haq, N.U., 2020. Responses of persimmon (Diospyros kaki) fruits to different fruit coatings during postharvest storage at ambient temperature. *Journal of Pure and Applied Agriculture*, 5(3), pp.26-32. <u>https://jpaa.aiou.edu.pk/wp-content/uploads/2020/10/JPAA\_2020\_5\_3\_26-32.pdf</u>
- Khan, T., Mazid, M., & Mohammad, F. (2011). A review of ascorbic acid potentialities against oxidative stress induced in plants. *Journal of Agrobiology*, 28(2), 97-111.
- Medlyn, B. E., Duursma, R. A., De Kauwe, M. G., & Prentice, I. C. (2013). The optimal stomatal response to atmospheric CO2 concentration: Alternative solutions, alternative interpretations. *Agricultural and Forest Meteorology*, 182, 200-203.
- Mehmood, A., Naveed, K., Ayub, Q., Alamri, S., Siddiqui, M.H., Wu, C., Wang, D., Saud, S., Banout, J., Danish, S. and Datta, R., 2021. Exploring the potential of moringa leaf extract as bio stimulant for improving yield and quality of black cumin oil. *Scientific Reports*, 11(1), pp.1-10. https://doi.org/10.1038/s41598-021-03617-w
- Mehmood, A., Naveed, K., Azeem, K., Khan, A., Ali, N. and Khan, S.M., 2018. 10. Sowing time and nitrogen application methods impact on production traits of Kalonji (Nigella sativa L.). *Pure and Applied Biology (PAB)*, 7(2), pp.476-485. http://dx.doi.org/10.19045/bspab.2018.70060
- Mehmood, A., Naveed, K., Jadoon, N., Ayub, Q., Hussain, M. and Hassaan, M., 2021.
  Phytochemical screening and antibacterial efficacy of black cumin (Nigella sativa L.) seeds. *FUUAST Journal of Biology*, *11*(1), pp.23-28.
  <u>https://fuuastjb.org/index.php/fuuastjb/article/download/592/433</u>

Mehmood, A., Naveed, K., Khan, S.U., Haq, N.U., Shokat, M.F., Iqbal, M., Ali, R., Nisar, S.,

Ahmad, J., Rehman, A.U. and Ur, S., Phytochemical screening, antioxidants properties and antibacterial efficacy of moringa leaves. *Journal of Xi'an Shiyou University, Natural Sciences Edition*, 18(10): 59-70. <u>https://www.xisdxjxsu.asia/V18I10-06.pdf</u>

- Mehmood, S., Ayub, Q., Khan, S.M., Arif, N., Khan, M.J., Mehmood, A., Shahzad, Q., ul Haq, N., Tanoli, M.T.Z. and Ayub, M.U., 2020. Responses of Fig Cuttings (Ficus Carica) to Different Sowing Dates and Potting Media under Agro-Climatic Conditions of Haripur. *RADS Journal of Biological Research & Applied Sciences*, *11*(2), pp.112-119. https://doi.org/10.37962/jbas.v11i2.268
- Mohamed, M., Mahmoud, H., Abo El-Fadle, H., & Abada, M. (2017). Response of superior grapevines to foliar sprayselenium with some vitamins. *Journal of Productivity and Development*, 22(2), 307-321.
- Moud, A. A. M., & Yamagishi, T. (2005). Application of projected pollen area response to drought stress to determine osmoregulation capability of different wheat (*Triticum aestivum* L.) cultivars. *International Journal of Agriculture and Biology*, 7(4), 604-605.
- Naeem, M., Bhatti, I. R. A. M., Ahmad, R. H., & Ashraf, M. Y. (2004). Effect of some growth hormones (GA3, IAA and Kinetin) on the morphology and early or delayed initiation of bud of lentil (*Lens culinaris Medik*). *Pakistan Journal of Botany*, *36*(4), 801-809.
- Rahbarian, R., Khavari-Nejad, R., Ganjeali, A., Bagheri, A., & Najafi, F. (2011). Drought stress effects on photosynthesis, chlorophyll fluorescence and water relations in tolerant and susceptible chickpea (*Cicer arietinum* L.) genotypes. *Acta Biologica Cracoviensia s. Botanica*.
- Rehman, A.U., Mehmood, A., Naveed, K., Haq, N.U., Ali, S., Ahmed, J., Rehman, S.U., Shoukat, M.F., Ayub, A., Usman, M. and Nisar, S., Integrated effect of nitrogen and sulphur levels on productive traits and quality of black cumin (Nigella Sativa L.). *Journal* of Xi'an Shiyou University, Natural Sciences Edition, 18(10): 38-58. https://www.xisdxjxsu.asia/viewarticle.php?aid=1269
- Sadak, M. S., Dawood, M. G., Bakry, B. A., & El-Karamany, M. F. (2013). Synergistic effect of indole acetic acid and kinetin on performance, some biochemical constituents and yield of faba bean plant grown under newly reclaimed sandy soil. *World Journal of Agricultural Sciences*, 9(4), 335-344.

- Saul, H., Madella, M., Fischer, A., Glykou, A., Hartz, S., & Craig, O. E. (2013). Phytoliths in pottery reveal the use of spice in European prehistoric cuisine. *PLoS One*, 8(8), e70583.
- Shah, S.U., Ayub, Q., Hussain, I., Khan, S.K., Ali, S., Khan, M.A., Haq, N., Mehmood, A., Khan, T. and Brahmi, N.C., 2021. Effect of different growing media on survival and growth of Grape (Vitus Vinifera) cuttings. *J Adv Nutri Sci Technol*, 1, pp.117-124.
- Shahzad, Q., Sammi, S., Mehmood, A., Naveed, K., Azeem, K., Ahmed Ayub, M.H., Hussain, M., Ayub, Q. and Shokat, O., 2020. 43. Phytochemical analysis and antimicrobial activity of adhatoda vasica leaves. *Pure and Applied Biology (PAB)*, 9(2), pp.1654-1661. <u>http://dx.doi.org/10.19045/bspab.2020.90174</u>
- Solaimalai, A., Sivakumar, C., Anbumani, S., Suresh, T., & Arulmurugan, K. (2001). Role of plant growth regulators in rice production–A review. *Agricultural Reviews*, 22(1), 33-40.
- Taran, S.N.U., Ali, S.A., Haq, N.U., Faraz, A., Ali, S. And Rahman, T.U., Antioxidant and antimicrobail activities, proximate analysis and nutrient composition of eight selected edible weeds of Peshawar region. *Journal of Xi'an Shiyou University, Natural Sciences Edition*, 18(9): 517-545. <u>https://www.xisdxjxsu.asia/viewarticle.php?aid=1224</u>
- Thomas, H., & Howarth, C. J. (2000). Five ways to stay green. *Journal of experimental botany*, *51*(suppl\_1), 329-337.