Pneumocystis Carnii Pneumonia Infection

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Abstract

By reading this chapter, readers will be able to understand what is pneumocystis carnii pneumonia (PCP) infection and how it affects immunocompromised patients and on prior treatment different diagnostic parameters are performed for sampling and afterwards treatment is recommended by a physican. Readers will be able to distinguish PCP patients infected with HIV/AIDS and PCP patients infected without HIV/AIDS. Also, by going through this chapter, readers will be able to distinguish between pneumonia, PCP fungal infection and the compatibility of COVID-19 with PCP fungal infection. Different treatment strategies are discussed in this chapter in order to treat PCP infected patients and risk factors are also considered so that by keeping in view these risk factors preventive measures are adopted to treat PCP fungal infected patients. Epidemiology of PCP infection is elaborated so that reader will be able to understand whether it is endemic or pandemic infectious disease. In addition, the pathophysiology of PCP is discussed so that reader should come to know its mode of transmission in lungs where this fungal infection starts replicating. Importantly, different case studies are discussed in this chapter for the purpose of understanding that more than one infection also affects PCP infected patients and different antibiotics therapy are performed along with adjunctive corticosteroids therapy for better recovey of the patient. It is

the expected responsibility of readers after studying this chapter to provide awareness of PCP infection among health care professionals and people.

1.1 History/Background

It is commonly called as Pneumocystis carini pneumonia (PCP pneumonia) / Pneumocystis jirovecii pneumonia (PJP). PCP is a fungal infection that can affect one or both of the lungs and its causative agent is pneumocystis jirovecii which is also called as Pneumocystic carini [1]. It mostly affects those individuals who are infected with HIV infections [2]. In 1980, this case was first reported in the USA and it occurred in an individual infected with HIV. In the beginning, scientists confused whether PCP is caused by protozoa or fungus so they classified it as protozoan but with the passage of time and advancement in medical knowledge they classified it as fungus [3]. Today, PJP is considered one of the several fungal infections which can cause severe life threatening problems in individuals suffering with AIDS/HIV infections [4]. In the USA, currently in the first decade of HIV epidemic 100,000 cases of PJP are reported but today due to the presence of anti-retroviral drug therapy (ART), individuals with HIV/AIDS are less likely to infect with PCP infection. PCP is a fungal infection that affects the lungs of individuals with weakened immune system and it is usually caused by Pneumocystis jirovecii. There are several reasons for weakened immune system such as cancer, HIV infection, AIDS, high dose corticosteroids or medicine taken after having bone marrow or organ transplant. This fungal infection mostly attacks in individuals suffering from HIV/AIDS infection but seems to rarely occur in healthy individuals who do not acquire AIDS/HIV infections [5]. Nearly, all the individuals suffering from PCP either have low oxygen levels in their blood (hypoxemia) at rest or an increase in their alveolararterial oxygen tension gradient which causes difficulty for the individual to breath [6].

1.2 Microbiology of PCP

PJP occurs in the respiratory tracts of humans and mammals and it belongs to unicellular fungi group. Distinct species flexibility is present between members of host specific genus. In 1909, Chagas first introduced this organism and then few years later Dr. Carni isolated it from infected rats and proposed this organism name as Pneumocystic carini and some years later Dr. Otto Jirovec and his group members found that this organism was also present in

humans, later, they isolated this organism and renamed it as Pneumocystis jirovecii. Because of that reason, this organism has dual names i.e. PCP and PJP. There was a clash among the scientists whether Pneumocystis jiroveci pneumonia belongs to protozoa group or trypanosome group, further nucleic acid biochemical analysis suggest that Pneumocystis RNA and mitochondrial DNA considered the organism as a fungus which is unicellular rather than a protozoa [7]. On further research, scientists found that these organisms exist in three structural forms such as trophozoite, sporozoite and cyst form. The trophozoite sometimes is called as trophic form due to its existence in clusters form. The sporozoite is also ranked as precystic form in which it lays down the resting phase, and the cyst is a kind of form containing several spores which are sometimes called as intracystic bodies.

1.3 Epidemiology of PCP

The PJP infection frequency began to rise more than 88% before the emergence of prophylaxis for PJP, but today with the advancement in antiretroviral drug therapy (ART) and prophylaxis, the frequency of PJP became decreasing i.e. 60-70% and it involves those individuals who may or may not be infected with HIV/AIDS [8]. PJP diagnosis is difficult because modern medical facilities are not present in under developed regions of the world. In Africa, its frequency is found to be occurred at the rate of 80% in infants suffering with pneumonia and other HIV infections [9]. In sub-saharan Africa, individuals suffering from tuberculosis, which is a serious lungs infections, also have a chance to be infected with PJP pneumonia [10]. Similarly in Pakistan PCP pneumonia prevalence rate is 16% and different antibiotic therapies are performed to prevent form PCP infection [11]. According to latest research prevalence rate of PCP infection seems to be 32-38% in India [10]. In USA, 75% overall PCP cases were reported [8] and in China PCP prevalence rate is 40% according to latest research [12]. In Malaysia, PCP prevalence rate is 60% and in Europe PCP detection ratio is 18% [13]. In Mozambique, PCP occurs at the incidence of 6.8% and a specimen study was performed via nasopharyngeal aspirates through PCR detection to confirm the infection.

In Malawi, 5% PCP detection ratio was observed and sample specimen was taken via lung aspiration through a PCR detection technique [8]. Similarly in Namibia, PCP detection ratio is 5%, but sample specimen was taken via sputum induction through Grocott's

methenamine silver (GMS) and PCR detection ratios [14]. In France, PCP occurs at the incidence of 26.1% and in Brazil 20% PCP detection rate was observed [8, 15]. In Uganda 4% PCP detection ratio was noticed and a specimen study was performed through bronchoalveolar lavage (BAL) test, and a modified Giemsa detection method was used in this regard [16]. In Vietnam, 3% PCP detection rate was seen and in Tanzania, 1.5% PCP detection ratio was observed and sample specimen was taken via oral route using a PCR detection technique [17]. In Poland, 21% PCP detection rate was observed, and in Malawi, PCP occurs at incidence rate of 9% and specimen study was performed via BAL test and detection methods used in this technique were indirect immunofluorescence technique (IFT) and PCP [18]. These are the different regions are of the world where different studies and techniques are performed for PCP detection ratio and by covering epidemiological factors of PCP fungal infection, it is concluded that PCP infectious diseases is pandemic disease because it is present in different regions of the world.

1.4 Etiology of PCP

The causative agent of PCP is the fungus pneumocystis jirovecii pneumonia [19]. Individuals with healthy immune system do not infect with this organism but those individuals who have weakened immune system may easily be affected by this fungus organism named as PCP. The immune system may be weakened due to several reasons such as in the case of cancer therapy, organ transplant and using those medicines (steroids) that suppress your immune system [20]. If PCP is not treated in proper way the patient's condition may get worsen. So, it is recommended to boost up your immune system in the case of this PCP infection.

1.5 Pathophysiology of PCP

Pneumocystis pneumonia is a worldwide infectious fungal disease and it occurs mostly in 3 to 4 years of children [21]. Furthermore, pre-clinical studies have suggested that PCP transmission is via airborne and clinical trials on humans has also been reported due to depressed immunity in individuals [22]. PCP occurs due to defective humoral and cellular immunity. Once PCP spores inhaled via alveoli the host organism starts replicating and ultimately causes disease. The role of immunity in the case of PCP infection is due to

following reasons such as 1) Defects in cellular or humoral immunity, 2) CD4+ production is low, 3) CD4+ T cell count is > 200cells/micro liter and 4) Development of PCP infection [9].

1.6 Risk Factors

It is most likely to develop in those individuals in which HIV/AIDS infection has been reported. It also occurs in individuals who are immunodeficient. In this regard, patients receive long term immunosuppressive therapy. It can also occurs in individuals who are at risk of malnutrition [22]. Patients whose organ transplantation is performed and they receive corticosteroids therapy and their immune system is suppressed that's why chances of PCP fungal infection began to increase. PCP fungal infection is more susceptible in those patients whose are hematologic and non-hematologic malignant including solid tumors and cancerous cells[23].

1.7 Pneumonia & Pneumocystis Carini Pneumonia

Pneumonia is an infection of lungs affecting one or both lung parts [24]. In this infection, alveoli are filled with fluid and pus. It causative agent is Streptococcus pneumoniae. Pneumonia affects mainly patients with 65 years of age or more and children about 2 years of age [25]. The signs and symptoms of pneumonia in adults are clearly observed but in case of children signs and symptoms are monitored carefully. They may have fever, cough or they may have difficulty in breathing and eating. Pneumonia classification is based according to infectious agent or place from where infection spread. Community acquired pneumonia is caused by viruses, bacteria or fungi. Hospital acquired pneumonia is caused when patient is admitted for another illness but after recovering from particular illness got infectious pneumonia disease and it is more severe because bacteria are more resistant to antibiotic therapies and patients who acquire this infection are already immunosuppressed. Hospital acquired pneumonia develops when people are visiting to attend the patient for longer time and it is also more resistant to antibiotics and can also caused by visiting out-patients clinics. Aspiration pneumonia is caused when patient inhaled food, drink, saliva or vomit and disturbs body's normal reflux mechanism and this can also be spread by excessive intake of alcohol.

Pneumocystis carini pneumonia is caused by fungal infection and can affect one or both lungs [26]. It is usually spread individuals who are either immunocompromised or have already be infected with AIDS or HIV. Before adopting treatment strategy, its diagnosis is performed based on PCP patient infected with or without PCP. Factors that increase the risk of PCP fungal infections include smoking, slcohol intake, dyspnea and malnutrition. From above discussion, the difference of pneumonia and PCP fungal infection is clearly observed. Further basic details of pneumonia and PCP fungal infection are given below to summarize our findings and by observing these findings, it can easily diagnose whether patient is infected with pneumonia or PCP fungal infection.

Pneumonia	Pneumocystis carini pneumonia	
Inflammation of lungs in one or both parts.	PCP is a fungal infection that may affect one or both parts of lungs.	
Alveoli are filled with fluid and pus.	Before adopting a treatment strategy, it is important to ensure that PCP patients are infected with only one causative agent, or more than one causative agent, or other causative agents are involved.	
Streptococcus pneumoniae is the most causative agent which causes inflammation in alveoli.	Its causative agent is pneumocysti jirovecii.	
Pneumonia develops when body immune system is weak and body fails sufficient organisms.	It lowers infected patient's immunous system and CD4+ level and different antibiotics along with adjunctive corticosteroids are given to treat PCH infection.	

Table 1.1 Difference between pneumonia and PCP

Factors that may lead to pneumonia are pre-existing lungs disease, recent influenza infections, smoking and upper respiratory tract infections may trigger	Factors that may lead to PCP are smoking, alcohol intake, and malnutrition. These conditions may trigger PCP infection.
pneumonia.	
Vaccines are available for treatment of pneumonia.	No vaccines are currently available for the treatment of PCP fungal infection.

1.8 COVID 19 and PCP Fungal Infection

Coronavirus disease 2019 (COVID-19) and pneumocystis pneumonia (PCP) have a lot of similarities and can be clinically indistinguishable at first in HIV-positive persons [27]. Diagnostic parameters are performed on dominant infection whether it is COVID-19 or PCP fungal infection. Similarities and differences based on COVID-19 and PCP fungal infection can be better described and explained with the help of case studies.

1.8.1 Case Study 1

During the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) pandemic, a 25year-old patient presented with significant hypoxemia despite using a non-rebreather mask. A massive right pneumothorax and severe interstitial illness were discovered on a chest X-ray. Despite the installation of a chest tube, hypoxemia persisted, prompting emergency intubation. A CT scan of the chest was performed, and the nasopharyngeal SARS-CoV-2 PCR was positive. His absolute CD4+ count was 32 cells/Mm³ and his HIV serology was positive. Because of his severe acquired immunodeficiency, radiographic results suggested a life-threatening co-infection with pneumocystis jirovecii, prompting therapy with trimethoprim–sulfamethoxazole, prednisolone, and remdesivir. Pneumocystis pneumonia (PCP) was confirmed four days later by bronchoscopic pneumocystis antigen. Clinically, the patient improved and was discharged from hospital successfully 21 days later.

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Figure 1.1 Depicting a chest X-ray and a CT scan of the chest at the time of presentation, (**A**) X-ray of the chest reveals a massive right pneumothorax and severe interstitial illness, (**B**) Coronal CT chest picture demonstrating apical cystic alterations, diffuse ground-glass opacities, thick consolidation, and pneumothorax, (**C**) An axial image of the most prominent apical cystic alterations and (**D**) Axial image with the chest tube revealing diffuse ground-glass opacities and a right pneumothorax .

1.8.1.1 Case Discussion

In both PCP and SARS-CoV-2 infection, widespread ground-glass nodules are the most common finding, making radiographic distinction challenging, especially in immunosuppressed patients. One-third of patients with serious PCP may develop cystic tumors [28, 29]. The identification of Pneumocystis jirovecii co-infection would have been difficult without these cystic radiographic features. As a result, in the present SARS-CoV-2 pandemic, knowledge of co-infections is vital in order to correctly diagnose and treat these co-infections, decreasing morbidity and mortality rates [30].

1.8.2 Case Study 2

With a three-week history of cough, myalgia, fever, and increasing dyspnea, a 54-year-old man was taken to a medical center. His BMI was normal, and he had an eight-year history of hypertension and type 2 diabetes as well as electrocardiographic signs of left ventricular hypertrophy. He had previously had two bouts of drug-sensitive pulmonary tuberculosis, both of which he had successfully treated. He was HIV positive at the time of admission, with a CD4+ count of 26 cells/L and a viral load of 2,447,646 copies/mL. His SARS-CoV-2 polymerase chain reaction (PCR) nasopharyngeal swab was likewise positive. He was moved to a field hospital for coronavirus illness (COVID), where he needed nasal prong oxygen to keep his oxygen saturation (SpO₂) at 96 percent. His oxygen needs increased within 24 hours of transfer, therefore he was transferred to our intensive care unit (ICU) for high-flow nasal cannula (HFNC) oxygen.



Figure 2.1 Radiograph of the right lower zone showing a severe deterioration

The above figure shows that on arrival, the patient's chest radiograph revealed bilateral ground-glass opacifications, mostly in the lower zone. He was started on empiric dexamethasone and therapeutic cotrimoxazole after a working diagnosis of COVID-19 and/or PCP. On the third day of hospitalization, however, persistent fever, haemodynamic instability, increased oxygen needs with worsening pulmonary infiltrates on chest X-ray, and growing inflammatory markers led to a diagnosis of nosocomial pneumonia. In the absence

of culture findings, empiric meropenem and fluconazole were started. The above figure shows the consolidation including all but the left upper zone was shown on this chest radiograph, indicating a severe deterioration. Sputum DFAT confirmed PCP, which was backed up by a beta-D-glucan level of > 500 pg/mL (normal 60). Acinetobacter baumannii was also found in blood cultures, for which colistin was started pending sensitivity. Legionella species were not found in the urine. On the eighth day after being admitted to the ICU, the patient died of increasing respiratory failure.

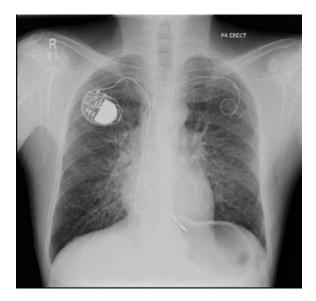


Figure 3.1 Radiograph of the left upper zone showing a severe deterioration.

1.8.2.1 Diagnosis

COVID-19 was initially regarded the most likely diagnosis in the emergency department, may be due to the relative frequency of COVID-19 during the peak of the epidemic and the resulting limitations in clinical reasoning. The diagnosis explored on the second presentation, 4 days later, were community acquired pneumonia (CAP) and moderate/severe COVID-19. Although the lymphopenia and bilateral chest X-ray alterations were consistent with COVID-19, the patient also had several unusual characteristics, such as oral thrush, weight loss, and a long clinical history. Following admission and a negative SARS-CoV-2 polymerase chain reaction test (PCR) result, Pneumocystis jirovecii pneumonia (PCP) was evaluated initially. A middle-aged man with shortness of breath, nocturnal sweats, and weight loss came to community care in England in March 2020. He was given oral antibiotics after a chest X-ray

revealed nothing unusual. He brought himself to emergency treatment a week later. In the absence of laboratory testing, he was discharged with a suspected mild COVID-19 diagnosis after a chest X-ray revealed bilateral apical ground glass alterations (Fig 1a). He was clinically stable without severe hypoxia and was thus discharged with a suspected light COVID-19 diagnosis. Four days later with substantial hypoxia, increasing bilateral upper lobe airspace shadowing on chest X-ray, anaemia (haemoglobin of 87 g/dL), lymphopenia (0.14 cells/Mm³), and increased C-reactive protein (212 mg/L), as well as yeast infections were observed.

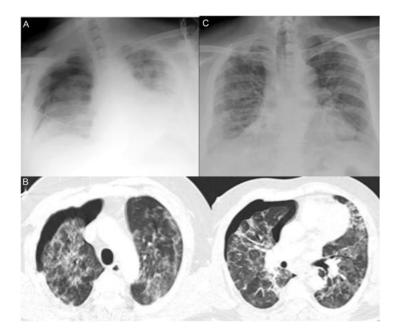


Figure 4.1 a) X-ray of the chest revealing bilateral apical ground glass alterations. **b)** An X-ray of the chest reveals increasing bilateral upper lobe airspace shadowing, **c)** and **d)** chest computed CT demonstrating extensive ground glass alterations with basal sparing [31].

1.8.2.2 Discussion

The patient's condition worsened on day 5 of hospitalization, and he was moved to the critical care unit for mechanical ventilation. SARS-CoV-2, influenza, parainfluenza, rhinovirus, adenovirus, respiratory syncytial virus, human metapneumovirus, and pneumocystis jirovecii were all found to be negative by PCR, and acid-fast bacilli (AFB)

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staining was negative. Co-trimoxazole was switched to IV pentamidine on admission day 9 due to bone marrow suppression and hyperkalemia. A bronchial alveolar lavage on day 10 revealed pneumocystis jirovecii confirming the diagnosis. CMV and herpes simplex type 1 (HSV-1) were also found in the lavage, but not SARS-CoV-2, respiratory viruses, or AFB. On day 15, a right tension pneumothorax occurred, as well as an intractable bronchopleural fistula. The patient died on the 17th day of his hospital stay, around 29 days after his first appearance in the community.

1.8.2.3 Conclusion

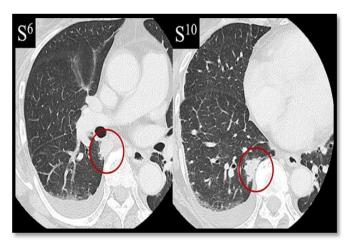
A lengthier clinical history previous to admission to emergency care is normal with COVID-19, oral thrush, and radiographic indications of apical alterations with basal sparing were among the clinical signals that may have prompted an earlier HIV test in the patient presented. Adding an HIV test to the order panel for all COVID-19 admissions, as well as educating and being attentive about potential differential illnesses like PCP, is one low-cost technique of enabling a differential diagnosis [32]. This will help determine how HIVpositive persons with and without COVID-19 should be treated, including cohort nursing and a focused therapy approach.

1.8.3 Case study 4

During prednisolone therapy for autoimmune hepatitis, a 76-year-old lady got pneumocystis pneumonia (PCP) (AIH). A haematological test done by her family physician revealed that the lady had excessive levels of hepatobiliary enzymes. She was sent to a hospital for more testing, and she was admitted to the hospital. She was diagnosed with autoimmune hepatitis (AIH) and began treatment on 40 mg of prednisolone per day. Her transaminase levels improved, and the quantity of prednisolone was lowered every two weeks. A fever was noted around the fourth week of medication. The fever reached 38°C in a few of days. Her chest X-ray revealed ground glass opacities, and she had cough also. An induced sputum cytodiagnosis was conducted since a respiratory illness was suspected. Her respiratory condition was followed after she was diagnosed with PCP.

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1.8.3.1 Discussion

Cotrimoxazole

[trimethoprim/sulfamethoxazole] was begun after the woman's fever was managed symptomatically. She had acute exhaustion, hyponatraemia, and a drop in platelets. ADRs were thought to be the cause of these signs and symptoms. Her cough went away once

her hyponatraemia was cured. Her fever subsided, and her pneumonia began to recover. Prednisolone was not lowered in dosage throughout this time and was kept at 15 mg/day for the whole 4-week period. Cotrimoxazole was kept on hand as a preventative measure, and AIH therapy was continued. As a result, multiple stomach ulcers and oral candidiasis was developed. She received treatment and was released on a daily dose of oral prednisolone of 10 mg. Her IgG and transaminase values were both in the normal range. She was taking prednisolone 10 mg every other day at the time of the previous follow-up.

1.8.4 Case study 5

On day 7 of the fifth chemotherapy session, a 69-year-old lady receiving biweekly pirarubicin hydrochloride, oncovin, cyclophosphamide, and prednisolone for mycosis fungoides developed a fever. The first chest CT pictures were unremarkable, and serum procalcitonin and -D-glucan

levels were normal. Meropenem and amphotericin B were used to treat a bacterial or fungal illness, but they were useless. A contrast-enhanced CT was performed around 10 days following the initial chest CT. Apart from two mass lesions in S6 and S10 in the right lower lobe, no source of fever was discovered.

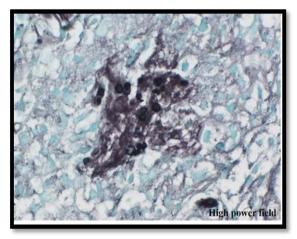


Figure 5.1 CT Scan revealing two masses of lesions for fever [33].

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In the right lung, pleural effusion was noted, which might be impacted by inflammation. Her SpO_2 level was within normal ranges, and she was asymptomatic. Bronchoscopy was conducted, and bronchial biopsy specimens stained with Grocott's staining indicated pneumocystis jirovecii cysts.

When compared to HIV-positive individuals, pneumocystis jirovecii is particularly difficult to identify in specimens from HIV-negative patients, who have a higher immune response to pneumocystis but fewer pathogens. Pneumocystis jirovecii can be detected using a polymerase chain reaction based on bronchoalveolar lavage fluid. Because the tumors were confined, biopsy and Grocott's staining proved to be the most crucial measures for a correct diagnosis in this case. Because the chest CT results were abnormal and the patient's blood -D-glucan level was within normal ranges, PCP was excluded from the differential diagnosis in this case. In a retrospective investigation of patients with PCP identified via bronchoalveolar lavage, Tasaka et al. found that blood -D-glucan level was the most accurate PCP predictor among serum levels of lactate dehydrogenase, -D-glucan, Krebs von den Lungen-6 (KL-6), and C-reactive protein. The threshold value for D-glucan concentration for PCP in the above-mentioned study was 31.1 pg/mL, with a sensitivity of 92.3 percent and a specificity of 86.1 percent. The reference range for -D-glucan, on the other hand, varies depending on the test technique used. Although the -D-glucan concentration was within normal ranges in this case, which might be due to the limited number of infections, there was insufficient data to evaluate if the D-glucan concentration represents the pneumocystis load in the lungs.

1.8.4.1 Conclusion

Depending on the patient's immunological state, PCP might appear with a variety of chest CT abnormalities. PCP is more frequent in immunocompromised persons and can be fatal. Even when chest CT images are abnormal, such as many lesions, PCP should be examined, and bronchoscopy should be attempted to diagnosis PCP if the patient's health allows. In suspected PCP cases, quick diagnosis and anti-PCP therapy are essential.

1.9 PCP Prognosis

The prognosis of PCP is worst due to its late diagnosis and it is major cause of death in US due to AIDS. It is most likely to occur in those patients who develop pneumothorax, and in this case patients receive mechanical ventilation. Currently, 20-50% cases have been reported due to large scale studies.

1.10 Clinical manifestations

These are basically signs and symptoms which distinguish and tells the medical expert whether infection is caused by Streptococcus pneumonaie and pneumocystis jirovecii. By keeping in view these findings diagnostic parameters are performed prior to treatment. PCP signs & symptoms and different findings are discussed in below section for further observations and understanding.

Symptoms	Signs	CXR Findings
Fever	Нурохіа	Diffuse, Bilateral, haz
Dyspnea	Tachypnea, tachycardia	Pneumothorax
Dry Cough	Inspiratory crackles	Pleural effusion, loba infiltrate, nodules les common
Pleuritic chest pain	Elevated A-a Gradient	CXR normal in 25%
Malaise	Chest exam normal in 50%	

Table 2.1 Signs and symptoms for diagnosis of PCP

1.11 Differential Diagnosis of PCP

Patients who are PCP infected may also be infected with other pneumonia diseases [34]. This condition is not necessary in this regard that PCP patient is infected with fungal disease only. PCP patient may also be infected with other fungal disease, that's why differential diagnosis is performed to ensure whether patient is infected with PCP or multiple viral, bacterial or fungal diseases are involved. Below table tell us about the differential diagnostic parameters of PCP infected patient.

Table 3.1 Differential diagnostic parameters of PCP infected patient

Acute respiratory distress syndrome (ARDS)	Cytomegalovirus	Lymphocytic Interstitial Pneumonia
Mycoplasma Infections	Viral pneumonia	Pulmonary embolism
Legionellosis	Tuberculosis	Mycobacterium avium Complex (MAC) Infection

1.12 Laboratory Studies

A lactic dehydrogenase (LDH) test is performed to detect the degree of lung injury [35]. Individuals who are infected with HIV are at great risk of PCP and it is uplifted in 90% patients. There exist an alternative to invasive testing procedure names as sputum P jirovecii PCR which is a time consuming method for sample collection and it is done in case of patient's respiratory failure. For PCP detection a sensitive test known as β -D-Glucan (BDG) is preferred which is comprised of Aspergillus, Candida and Pneumocystis but zygomycetes are excluded. The accuracy of this test is determined with Quantitative studies.

1.13 Chest Radiography

Those patients with defective immune system and other symptoms such as fever or respiratory signs are observed then in this regard chest radiography is performed. These results may be normal in patients with early mild disease. Common signs and symptoms include asymmetric infiltrates, pneumothorax, and pneumatoceles. In this overall process, a small amount of radiation is placed on the targeted organ such as lungs and then the image is detected in x-ray form.

1.14 Computed tomography

For the detection and imaging of PJP infection high-resolution computed tomography (HRCT) is used rather than CT scan because it can easily detect PJP patients infected with HIV infection. This process is monitored by the radiologist and after obtaining HRCT of the patients with the help of specified computers, concluded results are sent to the physician for treatment purposes.

1.15 Other Non-Invasive Tests

1.15.1 Pulmonary Function Tests

This test is performed on the basis of DLCO which stands for decreased diffusion capacity of carbon monoxide, patients who DLCO value is normal are less likely to be susceptible with PCP infection. Decreased DLCO value indicates higher risk of PCP infection (89-100%). When this test is compared with HRCT, then it is used to distinguish whether the PCP patient is infected with HIV/AIDS or the patient is infected with PCP infection only. In such type of tests, patient's observation is made on regular basis.

1.15.2 Pulse Oximetry

It should be calculated for all the patients at room temperature [36]. It should be calculated at rest or after some activity in specified patients. If hypoxemia is detected in which oxygen saturation is less than 90%, then arterial blood gas level should be attained along with corticosteroids.

1.15.3 HIV Testing

In case of possible HIV testing in PCP patients, observations and results made before the test and after the test should be evaluated carefully [37].

1.15.4 Laboratory Testing

In laboratory testing, blood sample of affected patient is evaluated according to LDH level.

1.16 Sputum Induction

Another method of detecting the PCP infection is via sputum induction [38]. In this process, the patient is nebulized with 3% hypertonic saline and specified patient is provided with box for sputum collection, after nebulization sputum is sent laboratory for detection of PCP infection caused by HIV or it is caused by only pneumonia. The sensitivity of this test is usually based on the effective technique applied in laboratory and its specificity varies from 99-100%.

1.17 Bronchoalveolar Lavage (BAL) Test

BAL test is performed if the sputum induction is negative and this test has higher diagnostic sensitivity, this test is performed on the recommendation of pulmonologist when the patient mental status is altered and the patient is unable to give sample by sputum induction. This test gives more sensitivity to detect PCP infection than sputum induction [39].

1.18 Lungs Biopsy

This test is performed to detect higher sensitivity and specificity when the results are obtained 100%. In this test, tissue samples are obtained from infected PCP patient's lungs for diagnostic purposes [40].

1.19 Histologic Findings

P Jiroveci organism cannot grow in vitro, therefore, histologic findings are observed before patient diagnosis. Several staining techniques are applied for PCP detection. 1) Crystal violet, 2) Giemsa, 3) Diff-Quik and 4) Wright stain [41].

For trophozoite and cyst form identification, mostly these staining methods are used.

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1.20 Treatment of PCP

Although PCP is declared as fungal pneumonia but it does not respond anti-fungal drugs. It is treated via TMP-SMX and other second line agents such as pentamidine and dapsone which are mostly along with pyrimethamine or atovaquone. Few successful case studies indicates that caspofungin can also treat PCP infection [42]. Treatment of PCP depends upon degree of illness and there are different parameters for tackling with PCP fungal infection which involves different drug combinations via oral or IV route depending upon the diagnosis and severity of the infection. There are two types of PCP patients, the first one are those who do not acquire HIV/AIDS infection and recovers within 4-5 days and the second one are those who acquire PCP infection and recovers within 21 days or longer [43].

1.20.1 Antibiotic Therapy

Table 4.1 Antibiotic Therapy for PCP

Mild to Moderate PCP (oral route)

First choice	Trimethoprim-sulfamethoxazole
Second choice	Trimethoprim (Proloprim) and dapsone
	or
	Clindamycin (Cleocin) and primaquine
Third choice	Atovaquone (Mepron)
Moderate to severe PCP (IV regimens)	
First choice	Trimethoprim-sulfamethoxazole
Second choice	Trimetrexate/leucovorin and oral dapson

or

Clindamycin (Cleocin phosphate) and

oral primaquine

Third choice

Pentamidine

1.20.2 Adjunctive corticosteroids therapy

In severe cases, corticosteroids therapy should be administered to the PCP patient because of suppression of immune response. Latest research indicates that patients who are infected with PCP fungal infection should not keep in contact with other immunocompromised patients.

Table 5.1 Adjunctive Corticosteroids Therapy for PCP

40 mg of prednisone twice daily
40 mg of prednisone once daily
20 mg of prednisone once daily

1.21 PCP Prevention

Smokers are at a greater risk of getting PCP infection, so it is advisable to quit smoking in order to prevent lungs from PCP infections. There is currently no vaccine available for PCP prevention. If a person is PCP infected, it is recommended to avoid direct contact with the person and follow safety precautions. If a patient immune system is weakened and CD4+ level is low then it is recommended to take medications that improves immune system and maintain CD4+ level. The medications should be taken only after physicians recommendations.

1.22 Conclusion

By reading this chapter, readers can easily get access about PCP infection and can easily distinguish between PCP patients infected with HIV or PCP patients infected without HIV. Hospitals should have separate wards for dealing with PCP patients for better patient care services and different tests are performed before diagnosis based on patient disease response and conditions. Antibiotics and adjunctive steroids are given to PCP patients for better recovery and maintaining their immune system. Patients must be observed carefully for clinical outcomes and it must also be assured that no drug interactions and toxicity occurs during treatment. If the treatment is carried out by a general family physician then he should be aware of this PCP fungal infection transmission and treatment protocols.

1.23 References

- 1. Sokulska M, Kicia M, Wesołowska M, Hendrich AB. Pneumocystis jirovecii—from a commensal to pathogen: Clinical and diagnostic review. Parasitology research 2015; 114(10): 3577-3585.
- 2. Lu J-J ,Lee C-H. Pneumocystis pneumonia. Journal of the Formosan Medical Association 2008; 107(11): 830-842.
- 3. Sritangratanakul S, Nuchprayoon S, Nuchprayoon I. Pneumocystis pneumonia: An update. J Med Assoc Thai 2004; 87(Suppl 2): S309-S317.
- 4. Arichi N, Kishikawa H, Mitsui Y, Kato T, Nishimura K, Tachikawa R, Tomii K, Shiina H, Igawa M, Ichikawa Y. *Cluster outbreak of pneumocystis pneumonia among kidney transplant patients within a single center*. in *Transplantation proceedings*. 2009. Elsevier.
- 5. Brown GD, Denning DW, Gow NA, Levitz SM, Netea MG, White TC. Hidden killers: Human fungal infections. Science translational medicine 2012; 4(165): 165rv13-165rv13.
- 6. White PL, Backx M, Barnes RA. Diagnosis and management of pneumocystis jirovecii infection. Expert review of anti-infective therapy 2017; 15(5): 435-447.
- 7. Bray PG, Barrett MP, Ward SA, de Koning HP. Pentamidine uptake and resistance in pathogenic protozoa: Past, present and future. Trends in parasitology 2003; 19(5): 232-239.
- 8. Morris A, Lundgren JD, Masur H, Walzer PD, Hanson DL, Frederick T, Huang L, Beard CB, Kaplan JE. Current epidemiology of pneumocystis pneumonia. Emerging infectious diseases 2004; 10(10): 1713.
- 9. Gingerich AD, Norris KA, Mousa JJ. Pneumocystis pneumonia: Immunity, vaccines and treatments. Pathogens 2021; 10(2): 236.

- 10. Rodríguez YDA, Wissmann G, Müller A, Pederiva M, Brum M, Brackmann R, De Paz VC, Calderón E. Pneumocystis jirovecii pneumonia in developing countries. Parasite: journal de la Société Française de Parasitologie 2011; 18(3): 219.
- 11. Zubairi ABS, Shahzad H, Zafar A. Clinical outcomes of pneumocystis pneumonia from a tertiary care centre in pakistan. JPMA. The Journal of the Pakistan Medical Association 2016; 66(11): 1367.
- 12. Chen M, Xu Y, Hong N, Yang Y, Lei W, Du L, Zhao J, Lei X, Xiong L, Cai L. Epidemiology of fungal infections in china. Frontiers of medicine 2018; 12(1): 58-75.
- 13. Velayuthan RD, Samudi C, Lakhbeer Singh HK, Ng KP, Shankar EM, Denning DW. Estimation of the burden of serious human fungal infections in malaysia. Journal of Fungi 2018; 4(1): 38.
- 14. Nowaseb V, Gaeb E, Fraczek MG, Richardson MD, Denning DW. Frequency of pneumocystis jirovecii in sputum from hiv and tb patients in namibia. The Journal of Infection in Developing Countries 2014; 8(03): 349-357.
- 15. Dunbar A, Schauwvlieghe A, Algoe S, Van Hellemond JJ, Reynders M, Vandecasteele S, Boelens J, Depuydt P, Rijnders B. Epidemiology of pneumocystis jirovecii pneumonia and (non-) use of prophylaxis. Frontiers in cellular and infection microbiology 2020; 10: 224.
- 16. Taylor SM, Meshnick SR, Worodria W, Andama A, Cattamanchi A, Davis JL, Yoo SD, Byanyima P, Kaswabuli S, Goodman CD. Low prevalence of pneumocystis pneumonia (pcp) but high prevalence of pneumocystis dihydropteroate synthase (dhps) gene mutations in hiv-infected persons in uganda. PLoS One 2012; 7(11): e49991.
- 17. Masoza TS, Rwezaula R, Msanga DR, Chami N, Kabirigi J, Ambrose E, Muro R, Mongella S, Hokororo A, Kwiyolecha E. Prevalence and outcome of hiv infected children admitted in a tertiary hospital in northern tanzania. BMC pediatrics 2022; 22(1): 1-9.
- 18. Gajewska M, Goryński P, Paradowska-Stankiewicz I, Lewtak K, Piotrowicz M, Urban E, Cianciara D, Wysocki MJ, Książek A, Izurieta P. Monitoring of community-acquired pneumonia hospitalisations before the introduction of pneumococcal conjugate vaccine into polish national immunisation programme (2009–2016): A nationwide retrospective database analysis. Vaccine 2020; 38(2): 194-201.
- 19. Schmidt JJ, Lueck C, Ziesing S, Stoll M, Haller H, Gottlieb J, Eder M, Welte T, Hoeper MM, Scherag A. Clinical course, treatment and outcome of pneumocystis pneumonia in immunocompromised adults: A retrospective analysis over 17 years. Critical Care 2018; 22(1): 1-9.
- 20. Suzuki T, Shimoda Y, Teruya K, Gatanaga H, Kikuchi Y, Oka S, Watanabe K. Case report: New development of fibrosing interstitial lung disease triggered by hiv-related pneumocystis pneumonia. BMC Pulmonary Medicine 2019; 19(1): 1-3.
- 21. García-Moreno J, Melendo-Pérez S, Martín-Gómez MT, Frick MA, Balcells-Ramírez J, Pujol-Jover M, Martín-Nalda A, Mendoza-Palomar N, Soler-Palacín P. Pneumocystis jirovecii pneumonia in children. A retrospective study in a single center over three decades. Enfermedades infecciosas y microbiologia clinica (English ed.) 2020; 38(3): 111-118.

- 22. Chesnay A, Paget C, Heuzé-Vourc'h N, Baranek T, Desoubeaux G. Pneumocystis pneumonia: Pitfalls and hindrances to establishing a reliable animal model. Journal of Fungi 2022; 8(2): 129.
- 23. Choo R, Naser NSH, Nadkarni NV, Anantham D. Utility of bronchoalveolar lavage in the management of immunocompromised patients presenting with lung infiltrates. BMC pulmonary medicine 2019; 19(1): 1-12.
- 24. Andrews CP, Coalson JJ, Smith JD, Johanson WG. Diagnosis of nosocomial bacterial pneumonia in acute, diffuse lung injury. Chest 1981; 80(3): 254-258.
- 25. Miller E, Andrews NJ, Waight PA, Slack MP, George RC. Herd immunity and serotype replacement 4 years after seven-valent pneumococcal conjugate vaccination in england and wales: An observational cohort study. The Lancet infectious diseases 2011; 11(10): 760-768.
- 26. Sjögren P, Nilsson E, Forsell M, Johansson O, Hoogstraate J. A systematic review of the preventive effect of oral hygiene on pneumonia and respiratory tract infection in elderly people in hospitals and nursing homes: Effect estimates and methodological quality of randomized controlled trials. Journal of the American Geriatrics Society 2008; 56(11): 2124-2130.
- 27. Xu X, Jiang X, Ma C, Du P, Li X, Lv S, Yu L, Ni Q, Chen Y, Su J. A deep learning system to screen novel coronavirus disease 2019 pneumonia. Engineering 2020; 6(10): 1122-1129.
- 28. Kanne JP, Yandow DR, Meyer CA. Pneumocystis jiroveci pneumonia: High-resolution ct findings in patients with and without hiv infection. AJR-American Journal of Roentgenology 2012; 198(6): W555.
- 29. Crans CA ,Boiselle PM. Imaging features of pneumocystis carinii pneumonia. Critical reviews in diagnostic imaging 1999; 40(4): 251-284.
- 30. Zahar J, Robin M, Azoulay E, Fieux F, Nitenberg G, Schlemmer B. Pneumocystis carinii pneumonia in critically ill patients with malignancy: A descriptive study. Clinical infectious diseases 2002; 35(8): 929-934.
- 31. Kelly S, Waters L, Cevik M, Collins S, Lewis J, Wu M-S, Blanchard TJ, Geretti AM. Pneumocystis pneumonia, a covid-19 mimic, reminds us of the importance of hiv testing in covid-19. Clinical Medicine 2020; 20(6): 590.
- 32. Stansell JD, Osmond DH, Charlebois E, Lavange L, Wallace JM, Alexander BV, Glassroth J, Kvale PA, Rosen MJ, Reichman LB. Predictors of pneumocystis carinii pneumonia in hiv-infected persons. Pulmonary complications of hiv infection study group. American journal of respiratory and critical care medicine 1997; 155(1): 60-66.
- 33. Kobayashi M, Tsubata Y, Shiratsuki Y, Hotta T, Isobe T. Multiple mass lesions in pneumocystis pneumonia. Cureus 2022; 14(1).
- 34. Sepkowitz KA. Pneumocystis carinii pneumonia in patients without aids. Clinical Infectious Diseases 1993; 17(Supplement_2): S416-S422.
- 35. Mossman BT, Marsh JP, Sesko A, Hill S, Shatos MA, Doherty J, Petruska J, Adler KB, Hemenway D, Mickey R. Inhibition of lung injury, inflammation, and interstitial

pulmonary fibrosis by polyethylene glycol-conjugated catalase in a rapid inhalation model of asbestosis. Am Rev Respir Dis 1990; 141(5 Pt 1): 1266-1271.

- 36. Selwyn PA, Pumerantz AS, Durante A, Alcabes PG, Gourevitch MN, Boiselle PG, Elmore JG. Clinical predictors of pneumocystis carinii pneumonia, bacterial pneumonia and tuberculosis in hiv-infected patients. Aids 1998; 12(8): 885-893.
- 37. Pulvirenti J, Herrera P, Venkataraman P, Ahmed N. Pneumocystis carinii pneumonia in hiv-infected patients in the haart era. AIDS patient care and STDs 2003; 17(6): 261-265.
- 38. Leigh T, Hume C, Gazzard B, Parsons P, Husain O, Collins J. Sputum induction for diagnosis of pneumocystis carinii pneumonia. The Lancet 1989; 334(8656): 205-206.
- 39. Golden JA, Hollander H, Stulbarg MS, Gamsu G. Bronchoalveolar lavage as the exclusive diagnostic modality for pneumocystis carinii pneumonia: A prospective study among patients with acquired immunodeficiency syndrome. Chest 1986; 90(1): 18-22.
- 40. Walzer PD, Perl DP, Krogstad DJ, RAWSON PG, SCHULTZ MG. Pneumocystis carinii pneumonia in the united states: Epidemiologic, diagnostic, and clinical features. Annals of internal medicine 1974; 80(1): 83-93.
- 41. Dako F, Kako B, Nirag J, Simpson S. High-resolution ct, histopathologic, and clinical features of granulomatous pneumocystis jiroveci pneumonia. Radiology case reports 2019; 14(6): 746-749.
- 42. Chen L, Zhang X, Peng X, Yang Y, Yu H. Baicalin tetrazole acts as anti-pneumocystis carinii pneumonia candidate in immunosuppressed rat model. Microbial pathogenesis 2019; 132: 59-65.
- 43. Dunphy L, Patel N, Palmer B, McKeown E. Missed opportunity to diagnose hiv with pneumocystis carinii pneumonia as its sequela. BMJ Case Reports CP 2020; 13(6): e235386.